

Today's Announcements

1. Test given back today. In general you did very well!
2. HW assigned today. Due Wednesday, 4/4/12.
3. Next Monday—1st discussion about Individual Projects.

Today's take-home lessons

(i.e. what you should be able to answer at end of lecture)

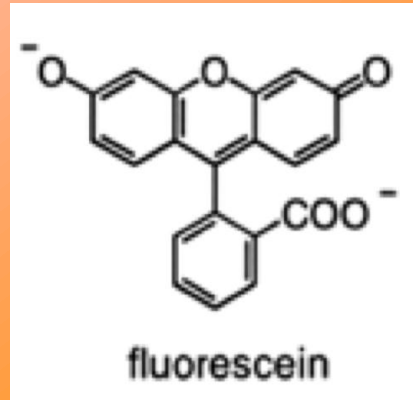
1. How to label things so they fluoresce—ext. labels, GFP.
2. Total Internal Reflection Fluorescence (TIRF)- near surface.
3. Accuracy and Resolution—how fine can you see.
4. Fluorescence. What is it (amplitude, time-scale)?

Fluorophores & Quantum Yield

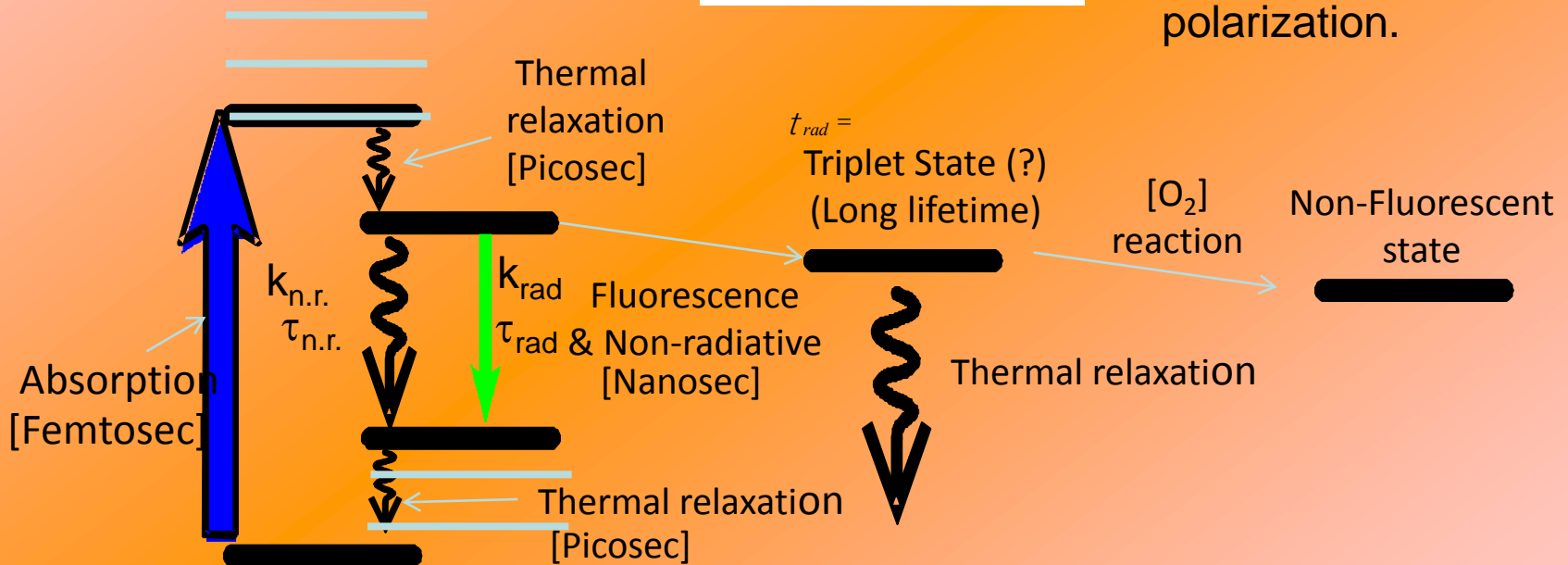
$$k = k_{\text{rad}} + k_{\text{n.r.}} ; \tau = \tau_{\text{rad}} + \tau_{\text{n.r.}}$$

$$\text{quantum yield} = k_{\text{rad}} / (k_{\text{rad}} + k_{\text{n.r.}})$$

Very good dyes have q.y. approaching one ($k_{\text{n.r.}} \approx 0$)



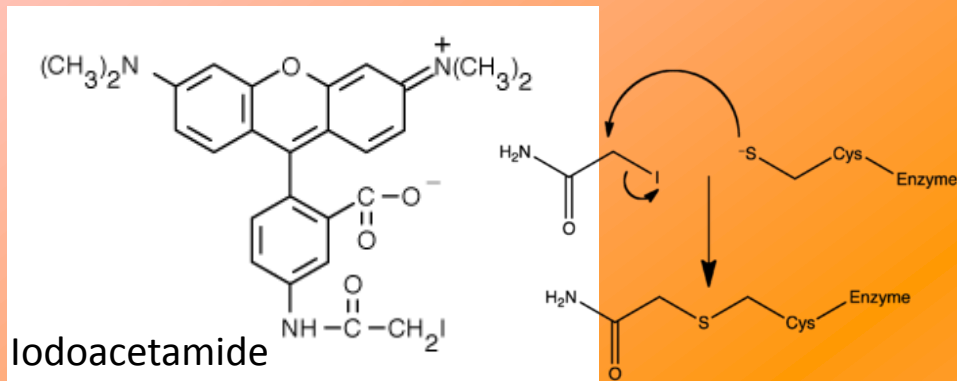
Have ≥ 1 electron that is free to move.
Excitation light moves e's around, i.e. a dipole, and it can re-radiate, often with polarization.



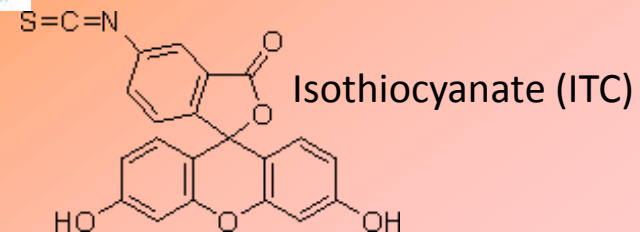
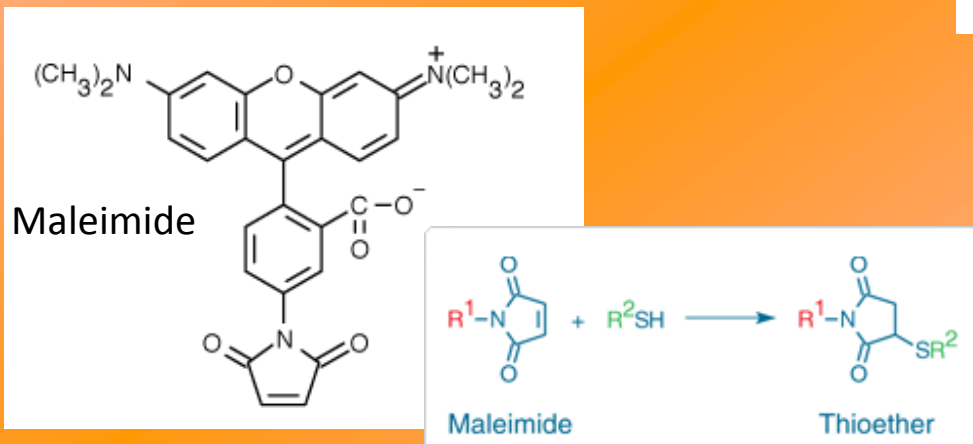
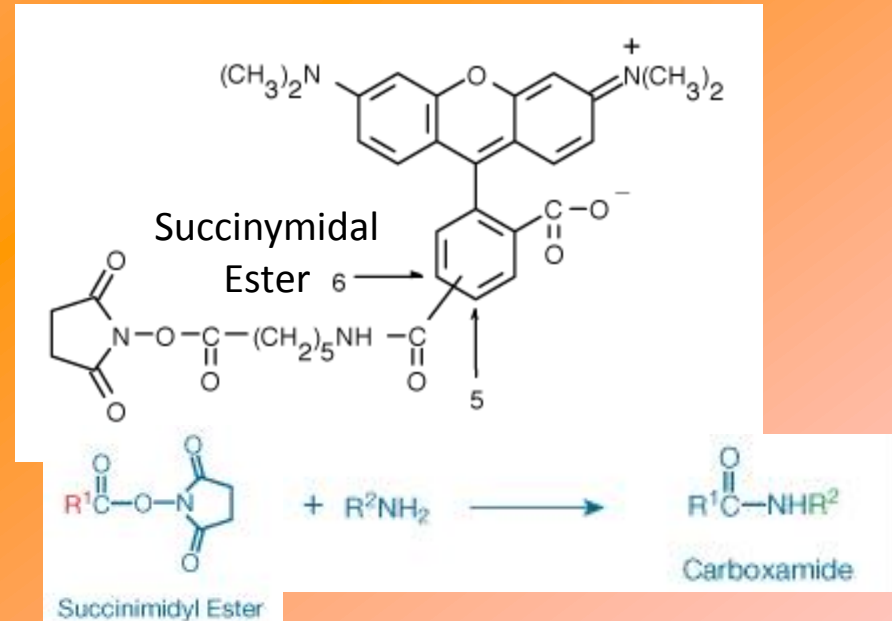
Basics of Labeling: In vitro

Bind to: free cysteines (R-SH) (often only one or a few in proteins)
: free lysines (R-NH₂) (many per protein)

Cysteine Reactive



Amine Reactive

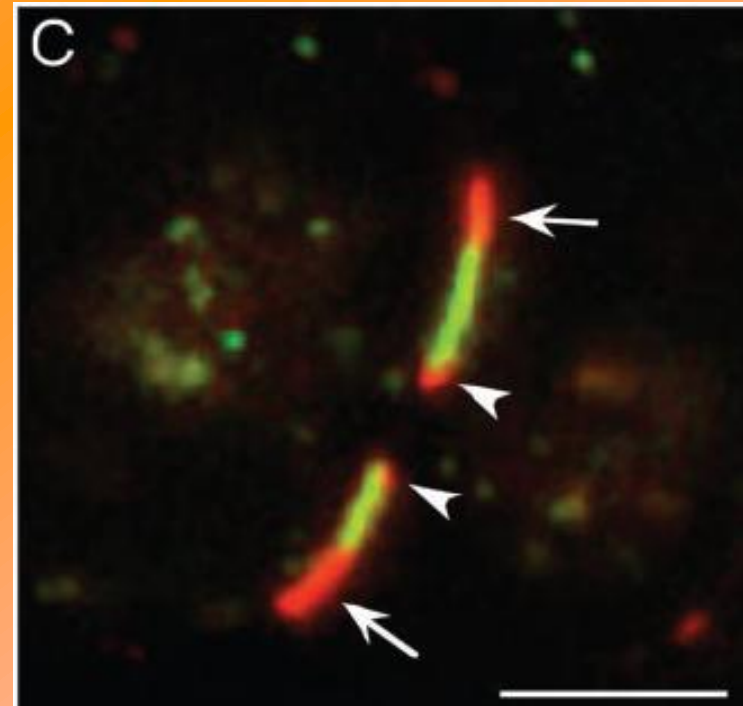
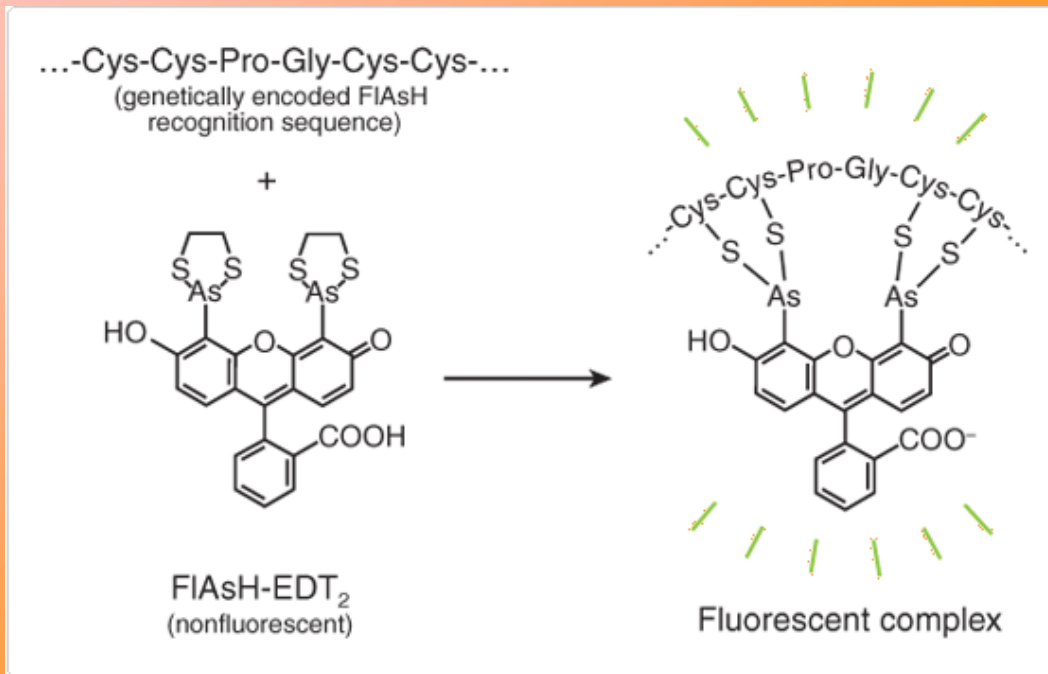


Basics of Labeling In vivo (inside cell)

Cell has a membrane, which is, in general, impermeant to dyes!

Bi-Arsenic FLASH, Fluorescent Proteins, SNAP-tag, Halo-tag

Bi-Arsenic FLASH, ReASH...

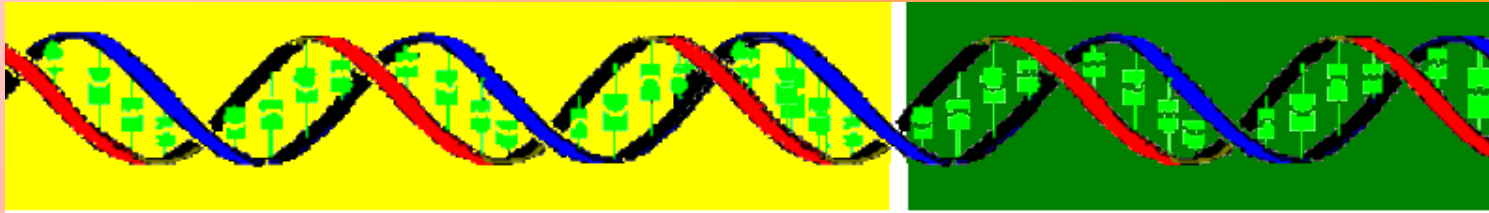


Tsien, Science, 1998

Tsien, Science, 2002

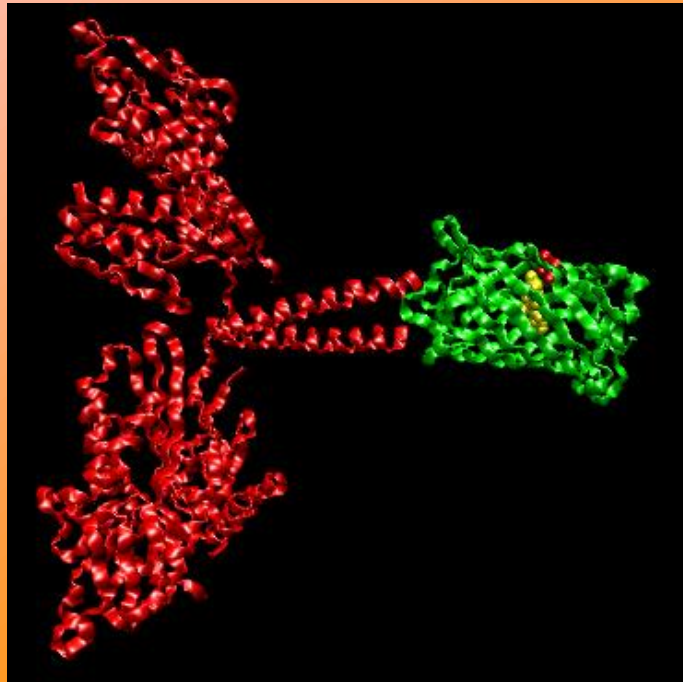
Green Fluorescent Protein (Nobel, 2009)

Genetically encoded dye (fluorescent protein)

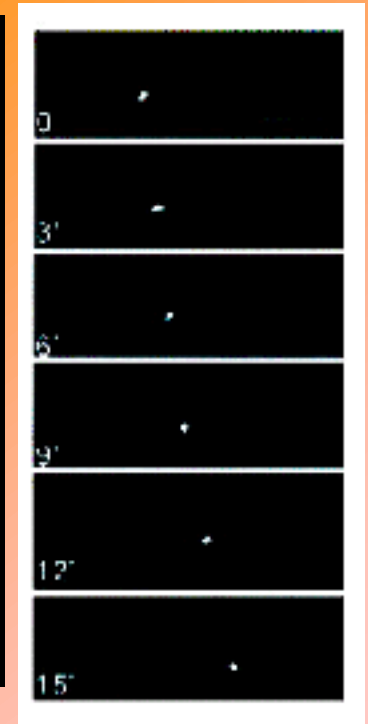


(Motor) protein

GFP



Kinesin – GFP fusion



Wong RM et al. PNAS, 2002

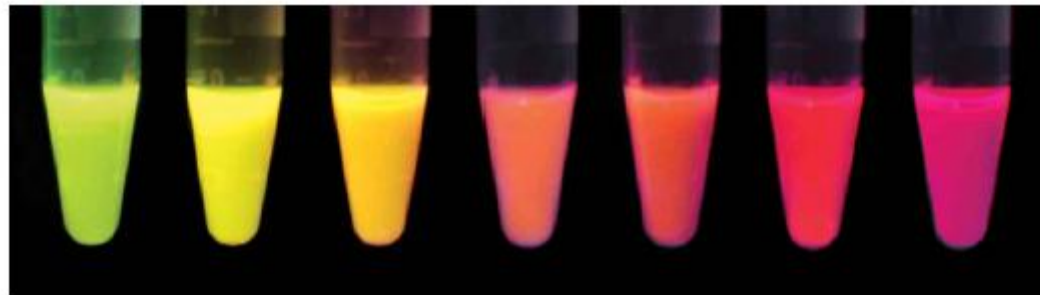
Genetically encoded → perfect specificity

Different Fluorescent Proteins

Absorption



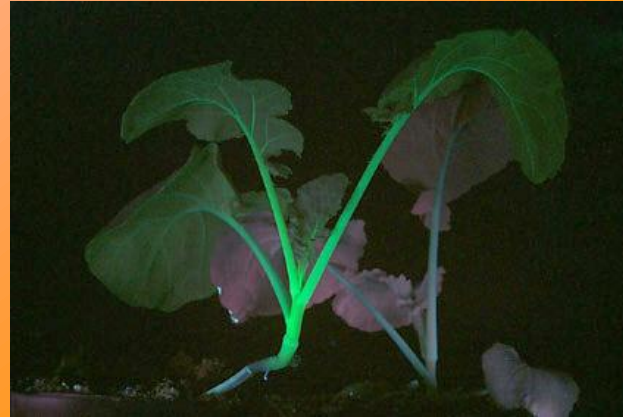
d



mHoneydew, mBanana, mOrange, tdTomato, mTangerine, mStrawberry, mCherry

Green Fluorescent Protein: Genetically-encoded dye

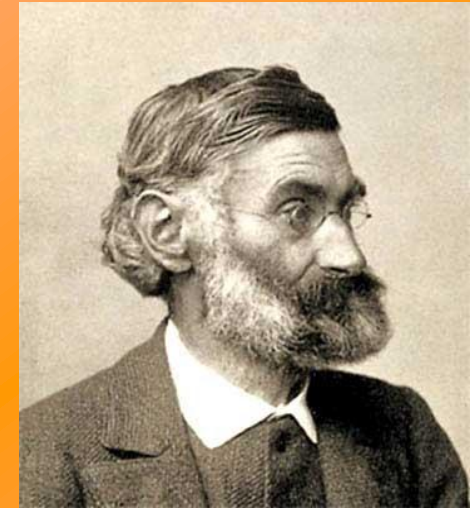
Fluorescent protein
from jelly fish



Aequorea victoria: the source of the fluorescent protein
Illustration by Jill Baker



How fine can you see? The Limits of Microscopy



Ernst Abbe

For visible microscopy,
Resolution is limited to ~250 nm

Ernst Abbe & Lord Rayleigh

Recent microscopy: 1-100 nm,

Here we present techniques which are able to get
super-accuracy (1.5 nm) and/or super-resolution (<10 nm, 35 nm)

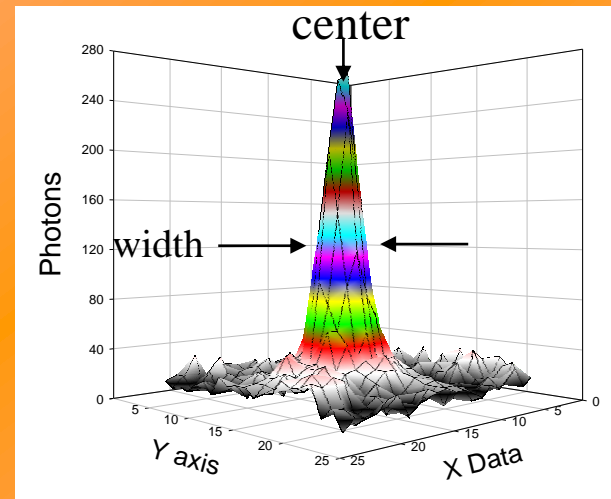
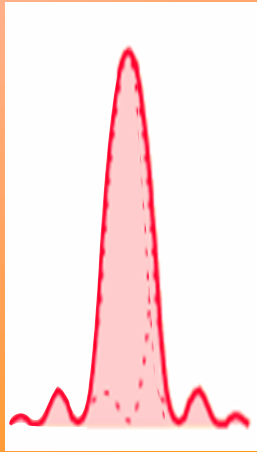
Super-Accuracy

(Accuracy \ll 250 nm: 1.5 nm, 1-500 ms)



FIONA

Fluorescence Imaging
with **One Nanometer**
Accuracy



Center can be found much more accurately than width

$$\begin{aligned} S/N &\approx \text{width} / \sqrt{N} \\ &\approx 250 / \sqrt{10^4} \approx 1.3 \text{ nm} \end{aligned}$$



When light gets dim,
what happens to your ability to find the center?

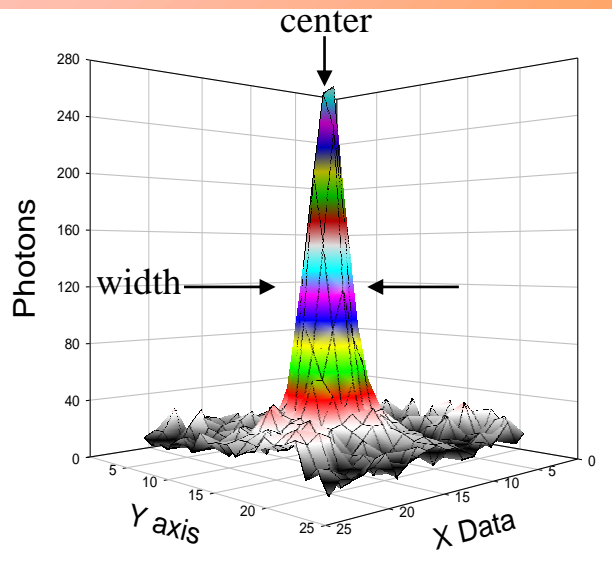
How well can you localize?

Depend on 3 things

1. # of Photons Detected (N)

2. Pixel size of Detector (a)

3. Noise (Background) of Detector (b)
(includes background fluorescence and detector noise)

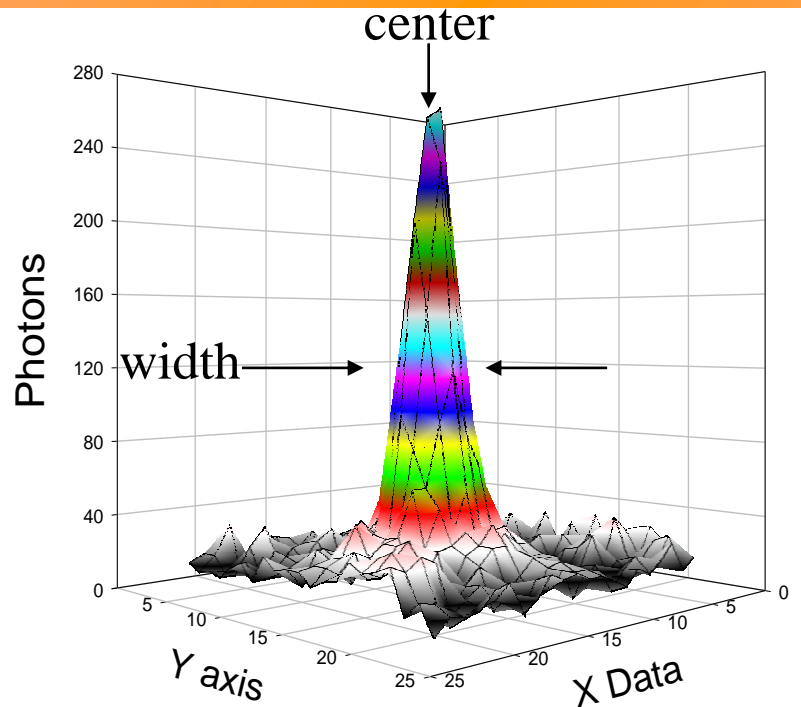
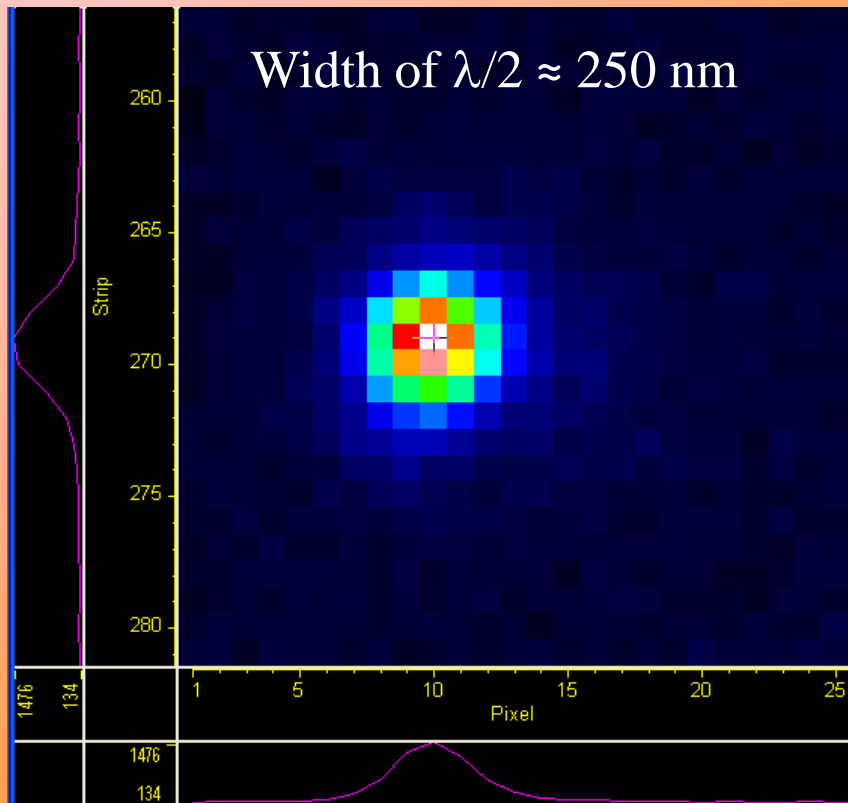


$$\sigma_{\mu_i} = \sqrt{\left(\frac{s_i^2}{N} + \frac{a^2/12}{N} + \frac{8\pi s_i^4 b^2}{a^2 N^2} \right)}$$

derived by Thompson et al. (Biophys. J.).

Diffraction limited spot: Single Molecule Sensitivity

$$\text{Accuracy of Center} = \text{width} / \text{S-N} \\ = 250 \text{ nm} / \sqrt{10^4} = 2.5 \text{ nm} = \pm 1.25 \text{ nm}$$



Enough photons (signal to noise)... **Center determined to ~1.3 nm**

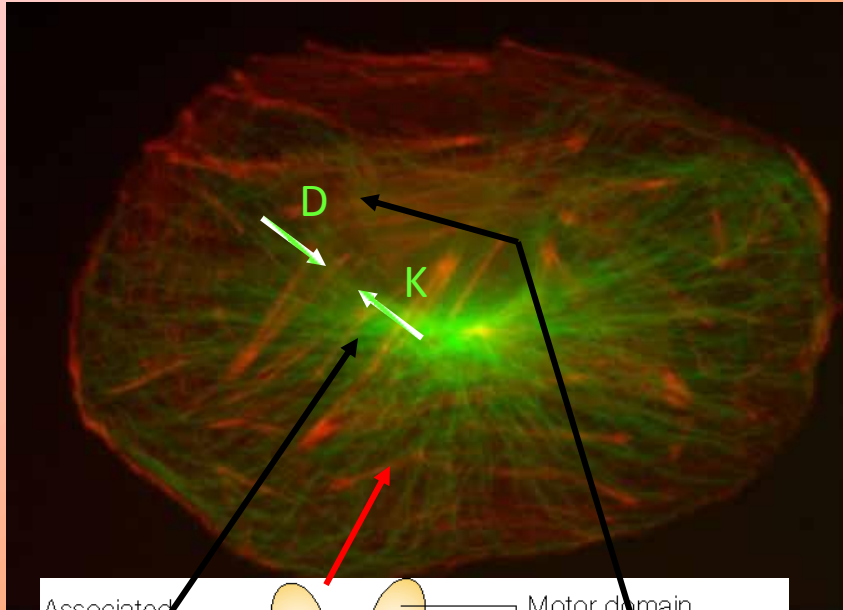
Dye lasts 5-10x longer -- typically ~30 sec- 1 min. (up to 4 min)

Start of high-accuracy single molecule microscopy

Thompson, BJ, 2002; Yildiz, Science, 2003

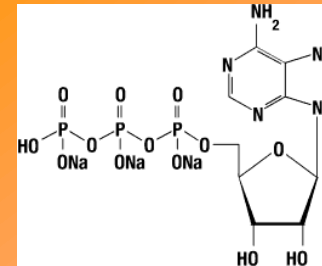
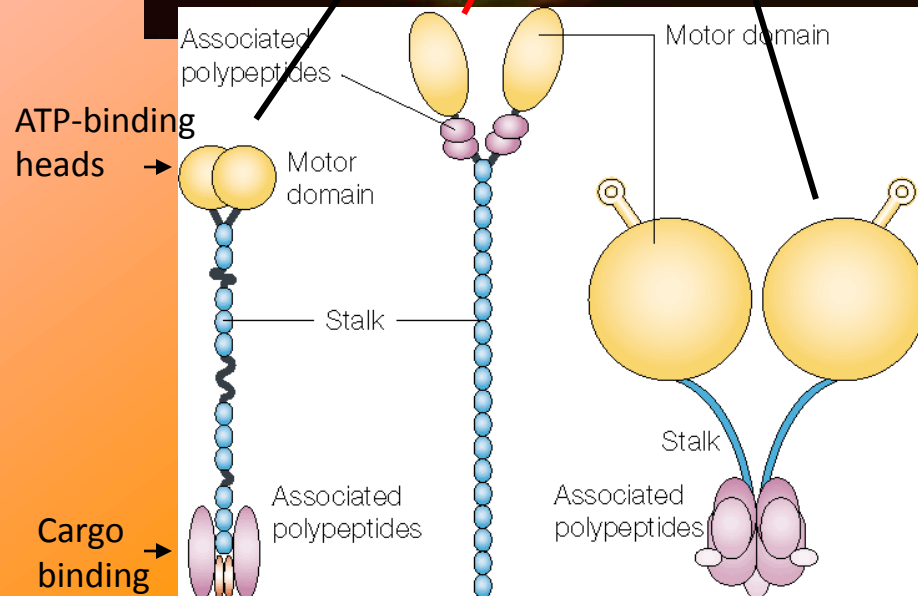
Biomolecular Motors: Intra- & Extra-Cellular Motion

Actin, μ tubules

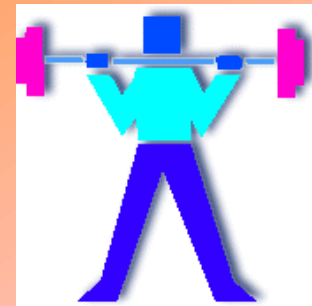


Characteristics

- nm scale
- Move along tracks
- intracellular directional movement
- cell shape changes & extracellular movement
- Use ATP as energy source



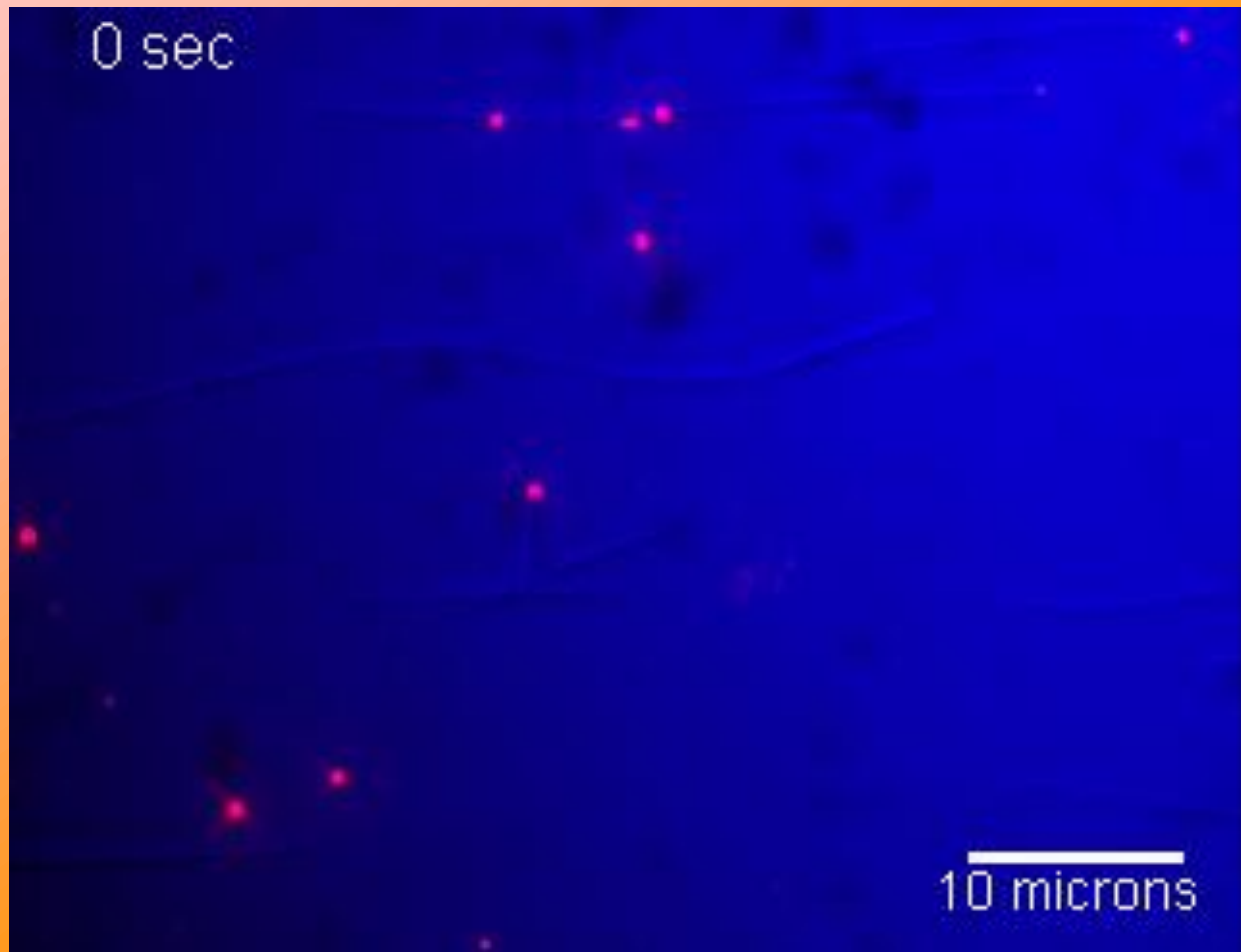
Nature Reviews



ATP \rightarrow mechanical work

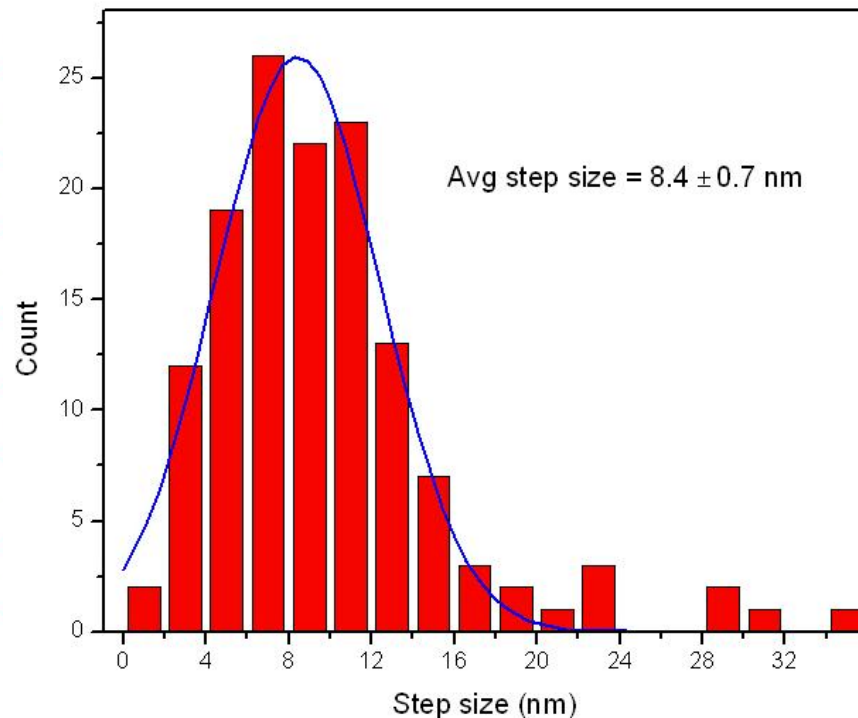
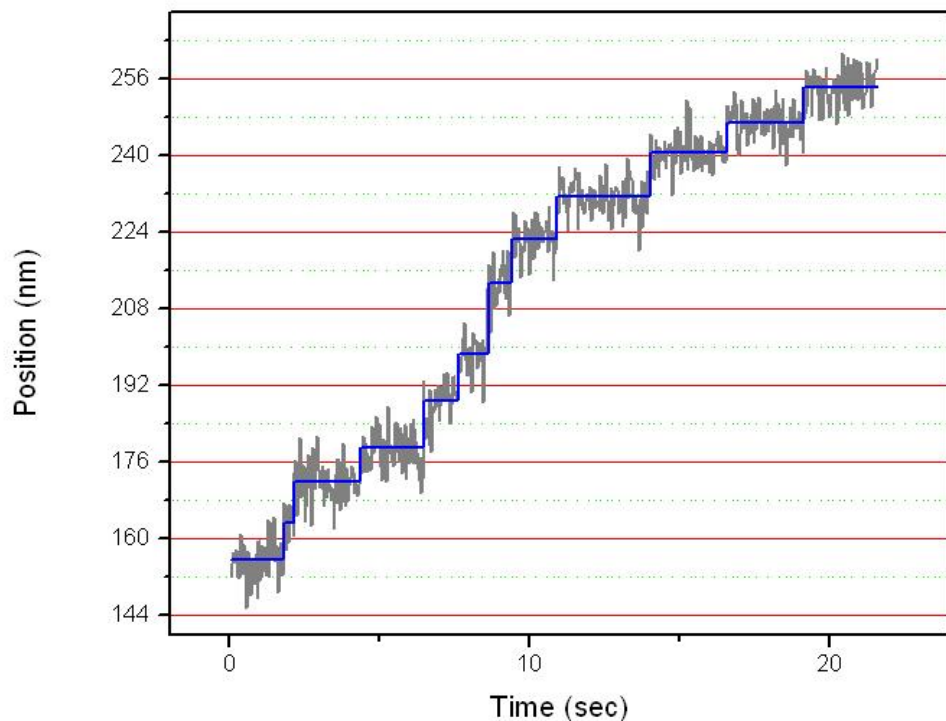
Kinesin	Myosin	Dynein	\leftarrow Motor
Microtubule	actin	Microtubule	\leftarrow polymer

Motility of quantum-dot labeled Kinesin (CENP-E)



8.3 nm/step from optical trap

Kinesin (Center-of-Mass) Moving

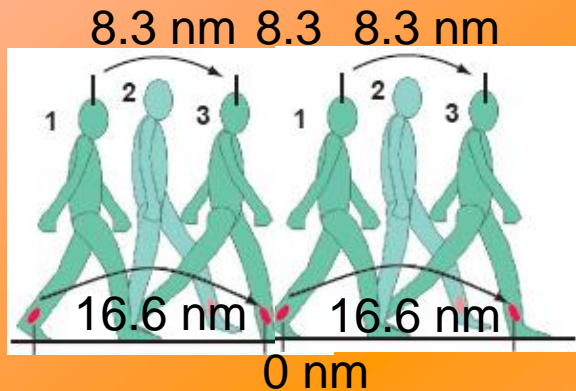
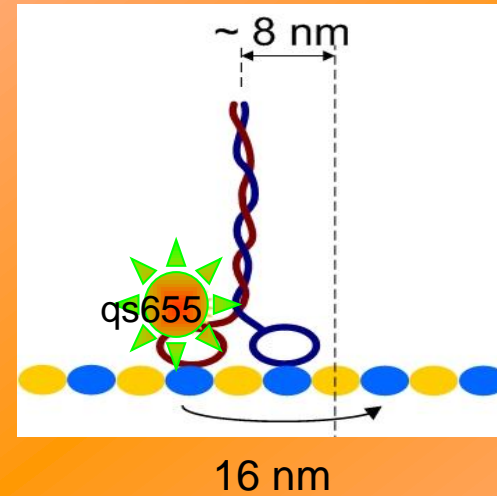


Kinesin moves with 8.4 nm /ATP step size.

Kinesin: Hand-over-hand or Inchworm?



8.3 nm, 8.3 nm

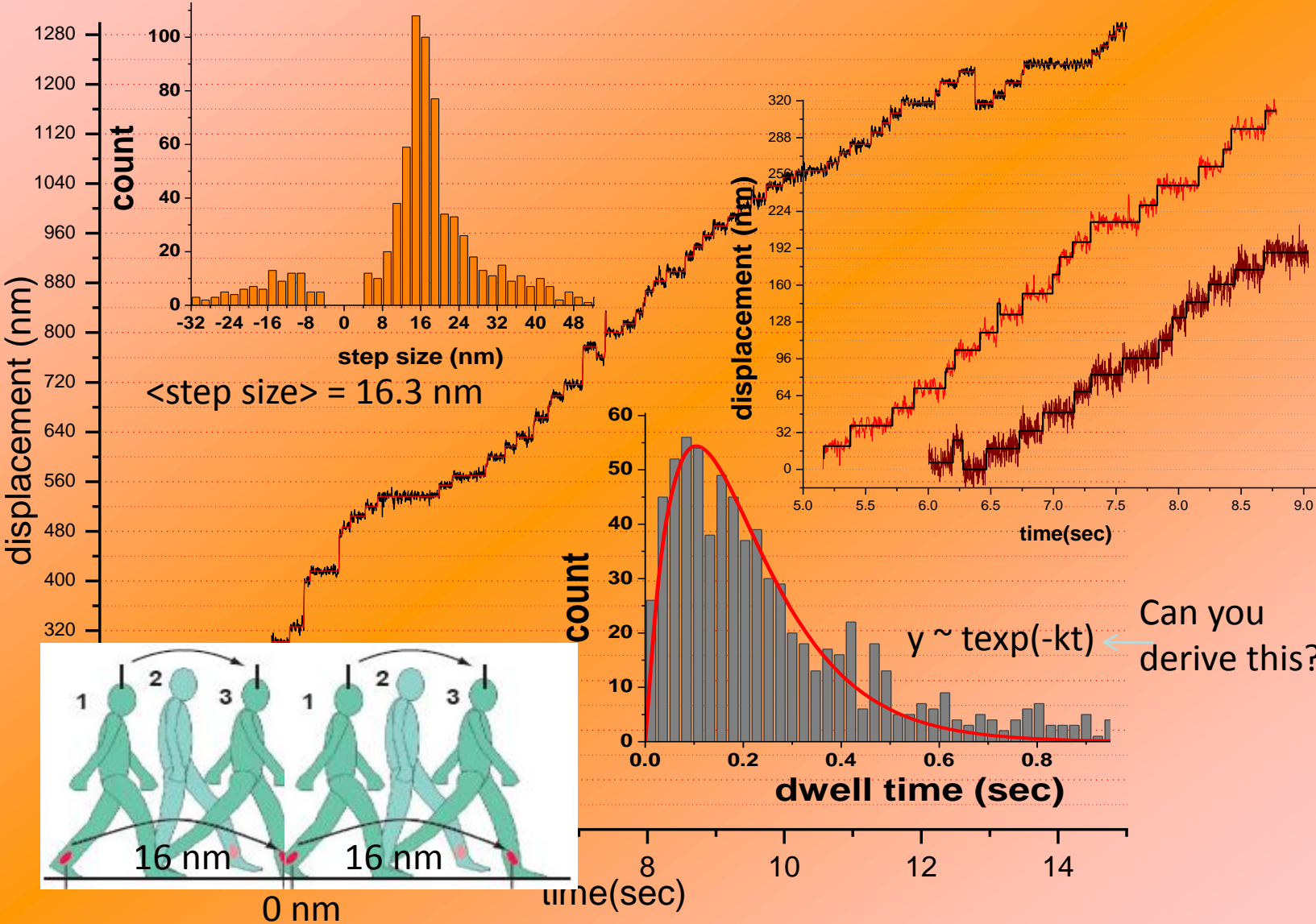


16.6, 0, 16.6 nm, 0...

[ATP] = 5 μ M ; 4 msec exposure time
 (Originally 0.3 μ M ; 500 msec exp. time)

pixel size is 160nm
 2 x real time

Kinesin



Can you derive this?

Takes 16 nm hand-over-hand steps (even at $5 \mu\text{M}$)

Class evaluation

1. What was the most interesting thing you learned in class today?
2. What are you confused about?
3. Related to today's subject, what would you like to know more about?
4. Any helpful comments.

Answer, and turn in at the end of class.