An LED and Spectroscopy System for Detecting Aflatoxin in Corn

Team 25 -- Jiahui Chen, and Foong Yee Wong ECE 445 Presentation -- Spring 2018 TA: Channing Philbrick

Objective

- Aflatoxins: toxic carcinogens produced by fungi, majorly Aspergillus flavus and Aspergillus parasiticus
- It grows in agricultural crops such as corn, peanuts, and tree nuts [1].
- Exposure to Aflatoxins can cause fatal damages to organs, especially for children. This may leads to stunted growth, liver damage, and liver cancer [2].
- We decide to create a low-cost Light Emitting Diodes (LEDs) spectroscopy system to let researchers from other places participate in this aflatoxin research.
- A group of researchers have built a device in the laboratory of Professor Matt Stasiewicz. We worked with John M. Hart to build and improve the reflectance spectroscopy system based on these principles for detecting the presence of aflatoxins, which can be reproduced and distributed.

Background

- Near infrared reflectance spectroscopy with a range of 550nm to 1700nm was used to evaluate the quality of grains and nuts [3]
- According to Ali Güneş's research, spectral data collected from NIR spectroscopy on a specimen, can be used to determine the presence of the toxins [4].
- Under ultraviolet light, the toxins exhibits Bright Greenish Yellow Fluorescence (BGYF) in visible light spectrum.
- The B-group Aflatoxins exhibit blue fluorescence (wavelength: 450 nm)
- The G-group exhibits yellow-green fluorescence (wavelength: 550 nm) under ultraviolet light [5].

Block Diagram

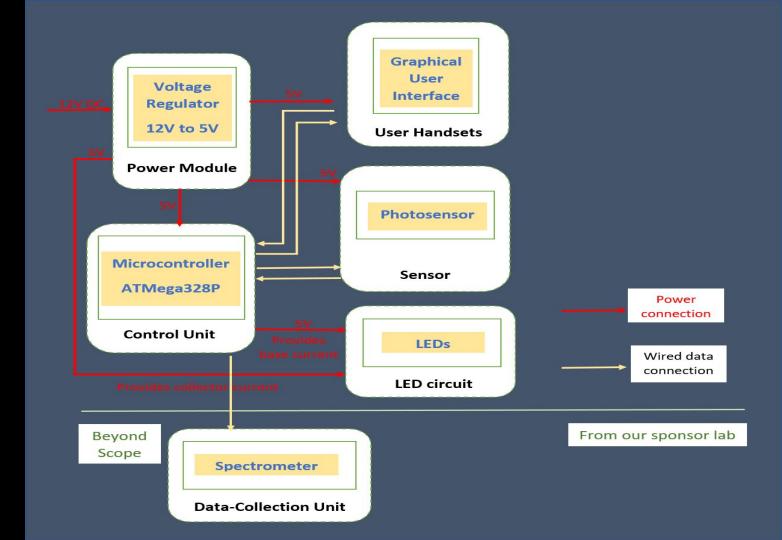


Figure 1: Block diagram

High Level Requirement

- Control the brightness of each individual groups of LED
- Voltage regulator can output 5V and provide sufficient current to the circuit
- Spectrum can be collected and displayed on computer/laptop

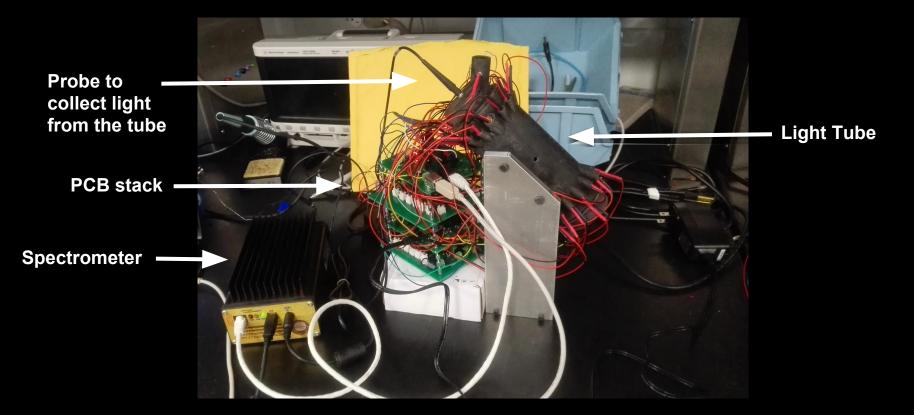
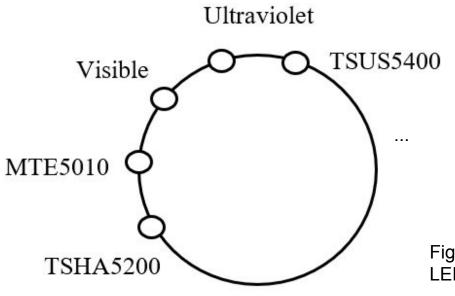


Figure 2: Set-up of the project

LED integration into tube assembly



...

Figure 3: Arrangement of LEDs around the light tube

Power Supply (12V, 5A)



| Figure 4: Power plugged |
|--------------------------|
| into socket on lab bench |

SPECIFICATION

| ORDER NO |). | SGA60U12-P1J | | |
|----------|------------------------------|--------------|--|--|
| | SAFETY MODEL NO. | SGA60U12 | | |
| | DC VOLTAGE Note.2 | 12V | | |
| | RATED CURRENT | 5A | | |
| | CURRENT RANGE | 0 ~ 5A | | |
| | RATED POWER (max.) | 60W | | |
| OUTPUT | RIPPLE & NOISE (max.) Note.3 | 80mVp-p | | |
| | VOLTAGE TOLERANCE Note.4 | ±3.0% | | |
| | LINE REGULATION Note.5 | ±1.0% | | |
| | LOAD REGULATION Note.6 | ±3.0% | | |
| | SETUP, RISE, HOLD UP TIME | 12ms/115VAC | | |
| | | | | |

Figure 5: Specification from the power adapter datasheet

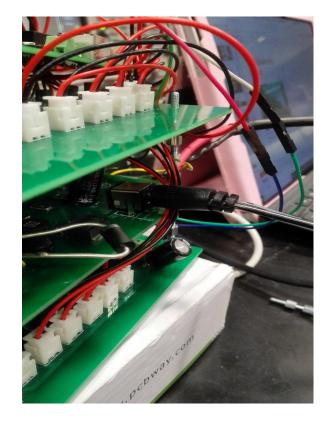


Figure 6: Power adapter connected to circuit board with a barrel connector

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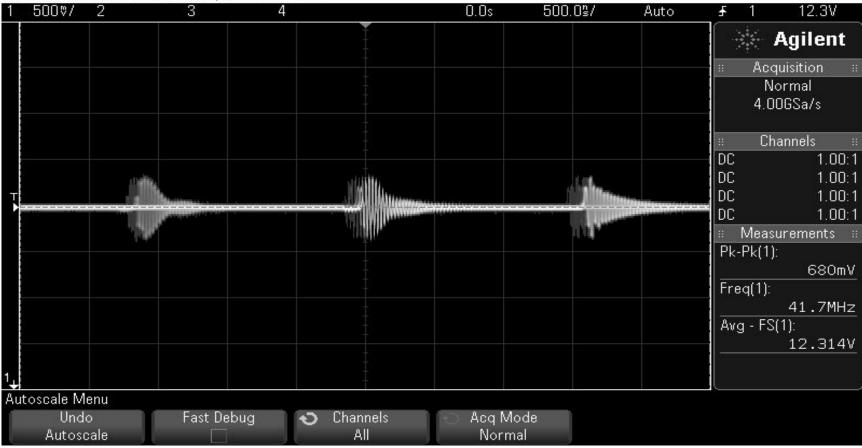


Figure 6: Screenshot captured from oscilloscope

Voltage regulator (LM2596-5.0)

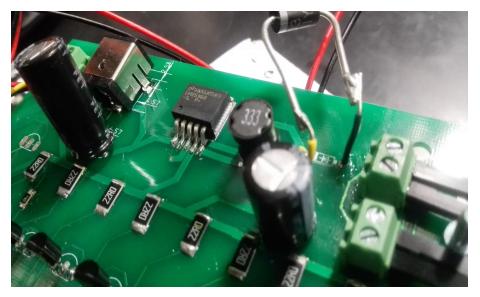


Figure 7: Voltage Regulator PCB

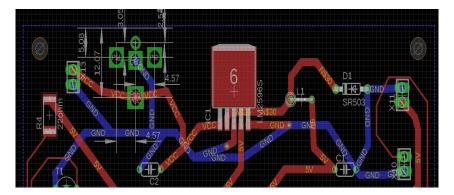


Figure 8: Voltage Regulator PCB Design

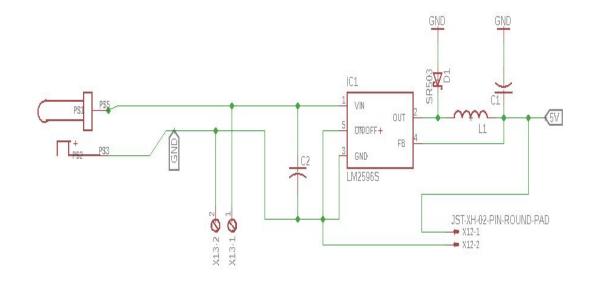
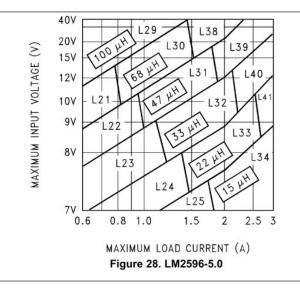


Figure 9: Voltage Regulator Schematic

 Input and output capacitor: Reduce noise

(For an aluminum electrolytic, the capacitor voltage rating must be approximately 1.5 times the maximum input voltage)[6]

- Inductor: Store energy and ensure the regulator to function well in continuous mode
- Schottky diode: prevent backflow of current



Voltage Regulator Datasheet [6]

9.2.1.2.3 Catch Diode Selection (D1)

 The catch diode current rating must be at least 1.3 times greater than the maximum load current. Also, if the power supply design must withstand a continuous output short, the diode must have a current rating equal to the maximum current limit of the LM2596. The most stressful condition for this diode is an overload or shorted output condition. See Table 4. In this example, a 5-A, 20-V, 1N5823 Schottky diode will provide the best performance, and will not be overstressed even for a shorted output.

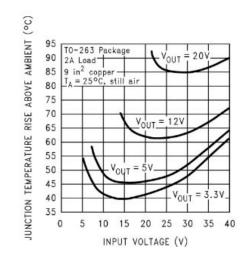
Table 4. Diode Selection Table

| | | 3-A D | IODES | | 4-A TO 6-A DIODES | | | | | | | |
|--------|----------|------------------------|----------|------------------------|-------------------|------------------------------|---------|------------------------|--|--|--|--|
| VR | SURFA | CE-MOUNT | THRO | DUGH-HOLE | SURF | ACE-MOUNT | THR | OUGH-HOLE | | | | |
| | SCHOTTKY | ULTRA FAST RECOVERY | SCHOTTKY | ULTRA FAST RECOVERY | скноттку | ULTRA FAST RECOVERY | сноттку | ULTRA FAST RECOVERY | | | | |
| | | All of | 1N5820 | All of | | All of | SR502 | All of | | | | |
| 20 V 🖇 | SK32 | these diodes | SR302 | these diodes | | these diodes | 1N5823 | these diodes | | | | |
| | are | | MBR320 | are | | are | SB520 | are | | | | |
| | 30WQ03 | rated to at least | 1N5821 | rated to at least | | rated to at least 50V. | | rated to at least | | | | |
| 30 V | SK33 | 50V. | MBR330 | 50V. | 50WQ03 | | SR503 | 50V. | | | | |
| | | | 31DQ03 | | | | 1N5824 | | | | | |
| | | | 1N5822 | | | | SB530 | | | | | |
| 40 V | SK34 | | SR304 | | 50WQ04 | | SR504 | | | | | |
| | MBRS340 | | MBR340 | | | | 1N5825 | | | | | |
| | 30WQ04 | MURS320 | 31DQ04 | MUR320 | | MURS620 | SB540 | MUR620 | | | | |
| 50 V | SK35 | 30WF10 | SR305 | | | 50WF10 | | HER601 | | | | |
| or | MBRS360 | | MBR350 | | 50WQ05 | | SB550 | | | | | |
| More | 30WQ05 | | 31DQ05 | | | | 50SQ080 | | | | | |

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Voltage Regulator Datasheet [6]



CIRCUIT DATA FOR TEMPERATURE RISE CURVE TO-263 PACKAGE (S)

| Capacitors | Surface-mount tantalum, molded D size | |
|------------|---|--|
| Inductor | Surface-mount, Pulse Engineering, 68 µH | |
| Diode | Surface-mount, 5-A 40-V, Schottky | |
| PCB | 9-square inch, single-sided, 2-oz. copper (0.0028") | |

Figure 41. Junction Temperature Rise, TO-263

Requirement and Verification (Voltage Regulator 1)

DOD V 2024A MM/E21024C2 Tup May 01 00/EC 40 2010

| 2.00V/ 2 | 3 | 4 | | 0.0s | 10.005/ | Auto? |) 5 1 | 4.99V |
|------------------|-----------|---|--------------|--|---------|-------|------------------|-------------------|
| | | | Ť | | | | | Agilent |
| | | | | | | | ii Ai | quisition |
| | | | | | | | | Normal 00GSa/s |
| | | | | | | | : (| hannels |
| | | | | | | | DC | 1.0 |
| | | | | | | | DC | 1.0 |
| | | | - | | | | DC | 1.0 |
| | | | Ţ. | | | | DC | 1.0 |
| | | | | | | | | surements |
| | | | | | | | Pk-Pk(| |
| | | | | | | | Eroa(1) | 300 |
| | | | | | | | Freq(1) | i. Low sig |
| | | | | | | | Avg - F | |
| | | | | | | | , rug i | 4.974 |
| | | | | | | | | |
| | | | | | | | • | |
| ear Measurements | | | | | | | | |
| Clear Meas 1 | Clear Mea | | Clear Meas 3 | Clear Mea | | Clear | | |
| Pk-Pk(1) | Freq(1) | | Avg - FS(1) | <none< td=""><td>></td><td>All</td><td></td><td></td></none<> | > | All | | |

Figure 10: Voltage Regulator 1 Output

Requirement and Verification (Voltage Regulator 2)

| 2.00V/ | 2 | 3 | 4 | | 0.0s | 10.00 % / | Auto? |) 1 1 | 4.99V |
|------------|-----------|---------|-------|--------------|--|------------------|-------|----------------------|-------------------------------------|
| | | | | | | | | | Agilent |
| | | | | | | | | N | quisition ormal IOGSa/s |
| | | | | | | | | III CH DC DC | annels 1.00: 1.00: |
| | | | | | | | | DC DC | 1.00: 1.00: 1.00: urements |
| | | | | | | | | Pk-Pk(1) Freq(1): | |
| | | | | | | | | Avg - FS | Low sign: (1): 4.9594 |
| lear Measu | rements M | enu | | | | | | | |
| Clear M | | Clear M | eas 2 | Clear Meas 3 | Clear Me | as 4 | Clear | | |
| Pk-Pk | | Freq | | Avg - FS(1) | <none< td=""><td></td><td>All</td><td></td><td></td></none<> | | All | | |

Figure 11: Voltage Regulator 2 Output

LED Selection

TSUS5400 950nm

TSHA5200 870nm

MTE5010 1050nm

MTE3650 365nm

C513A 450nm, 600nm

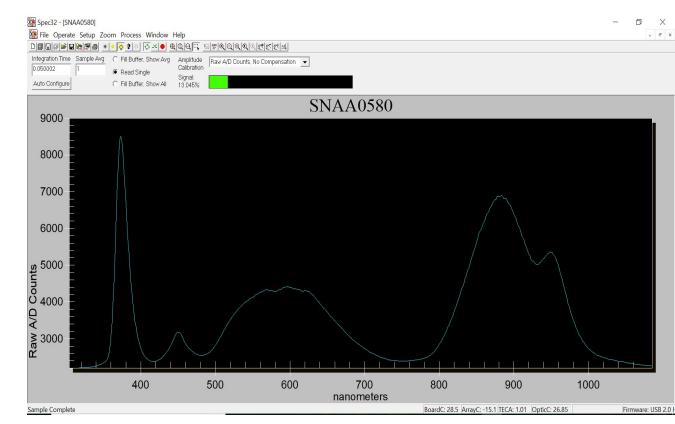


Figure 12: Spectrum of LEDs used in experiment

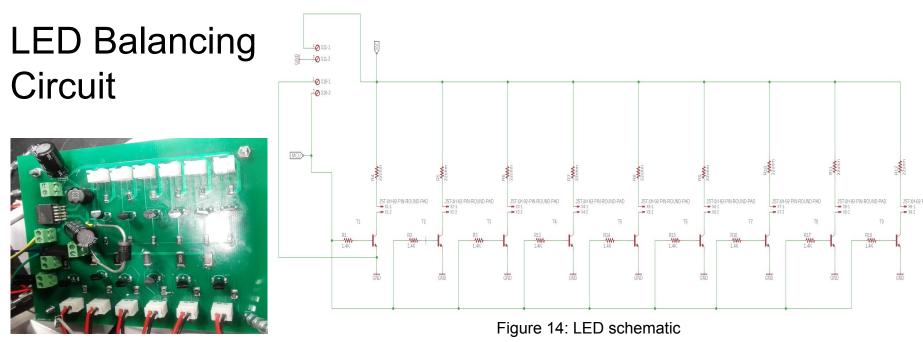


Figure 13: LED PCB

$$V_{CE} = 0.3V$$
 $V_{BE} = 0.6V-1.0V$
 $h_{FE} = 35-50$ $I_{B} \le 40mA$

$$R_c = \frac{V_{cc} - V_{led} - V_{CE}}{I_c} \qquad I_B$$

$$R_{B}= rac{V_{pin}-V_{BE}}{I_{B}}$$

 $\frac{I_c}{h_{FE}}$

Maximum Brightness

| LED part number | Forward Voltage from Datasheet (V) | Actual voltage across the LED (V) |
|-----------------|---------------------------------------|--|
| TSUS5400 | 1.3 | 1.385, 1.365, 1.394, 1.428, 1.358, 1.585 |
| TSHA5200 | 1.5 | 1.401, 1.411, 1.387, 1.386, 1.398, 1.385 |
| MTE5010 | 1.2 | 1.236, 1.230, 1.237, 1.228, 1.229, 1.226 |
| MTE3650 | 3.5 | 3.352, 3.389, 3.360, 3.389, 3.344, 3.348 |
| C513A | 3.2 | 3.124, 3.092, 4.85, 3.036, 3.080, 3.082 |

LED and Sensor

MTE1077N1-R:

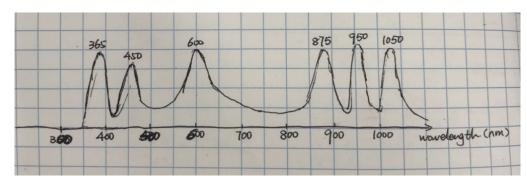


Figure 15: Wavelength of LEDs used in experiment

SD5600 honeywell sensor:

770nm visible light LED

(peak wavelength: 850, relative sensitivity at 770 nm: 0.9)

Light is detected: ~1

Light is blocked: ~1000

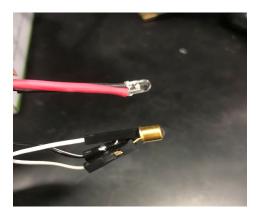


Figure 16: 770 nm LED and SD5600 sensor

| 💿 СОМЗ | | | | | | | <u>2</u> | - 🗆 | Х | 💿 СОМЗ | | | | | | | - | - 🛛 | Х |
|------------|------|------|------|------|------|----------------|-------------|----------|-----------|------------|---|---|---|---|---|--------------|---------------------|-------|-----------|
| | | | | | | | | | Send | | | | | | | | | | Send |
| 1017 | 1017 | 1017 | 1016 | 1017 | 1017 | 1017 | 1016 | 1017 | 1 ^ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | |
| | | | | | | | | | × | | | | | | | | | | × |
| < | | | | | | | | | > | < | | | | | | | | | > |
| Autoscroll | | | | | | No line ending | ✓ 9600 bauc | l 🗸 Clea | ar output | Autoscroll | | | | | | No line endi | ng \sim 9600 baud | ∨ Cle | ar output |

Figure 17: sensor reading: When light is blocked

Figure 18: sensor reading: when light is detected

Voltage across 770 nm LEDs: 1.666V, 1.680V (forward voltage is 1.55V).

Controller (FT232RL-Atmega328P)

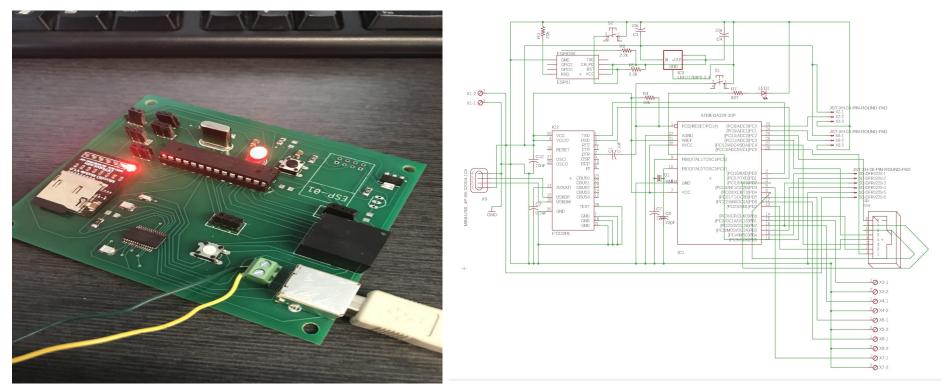


Figure 19: Controller PCB

Figure 20: Controller schematic

Flow Chart for programming

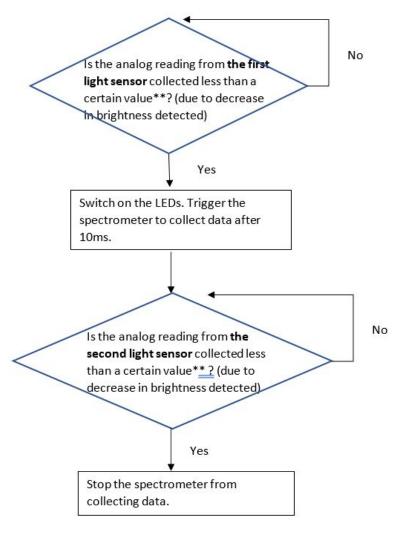


Figure 21: Flowchart to program light sensor, LEDs and spectrometer

Steps to configure the spectrometer:

- 1. Adjust the integration time to 0.005s
- Change the amplitude calibration to "Raw A/D Counts, no compensation"
- Click on 'Setup' and choose 'Trigger Mode', choose "Fast Asynchronous Clocking" and "External" (TTL input signal) for trigger control

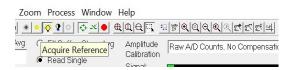


| Area Array Trigger Mode | × |
|---|---|
| Detector Clocking Mode C Synchronous Clocking (Auto M Fast Asynchronous Clocking () | |
| ✓ Bin All Rows (Highest Speed) | 🗹 Binning 🔲 Clamp |
| Trigger Control C Internal (Self-triggering) C External (TTL input signal) | Buffer Control Read Single Fill Buffer, Return Average Fill Buffer, Return All |

Steps to configure the spectrometer:

- 4. Collect the background brightness (with lights off)
- 5. Collect the reference brightness (with lights on)
- 6. Setup the trigger mode back to synchronous clocking and internal (self-triggering) for trigger control
- 7. Click on "Scan Continuous"

| File Operate | e Setup Zoom Process Window | Help |
|---------------------------|----------------------------------|--------------|
| | 號閉욜 ☀ ◦ Її ♀ ⊂ ♀ ● € | ŧ∣ℚ∣∈ |
| Integration Time 0.005002 | Sample Avg Avg Read Single | Amp Calib |
| Auto Configure | C Fill Buffer, Show All | Sign 3.37 |



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|---------|---|---|---|
| 15 🔹 | <mark> ● ≍ ●</mark> ⊕ ≍ ● € | A A A | ₽ ₩ € @ Q @ 0 |
| ole Avg | Fill Buffer, Scan Con Read Single Fill Buffer, Show All | ntinuous Campration Signal: 3.374% | Raw A/D Counts, |

Figure 23: Spectrometer software

PWM (pin 5, 11, 12, 15, 16, 17)

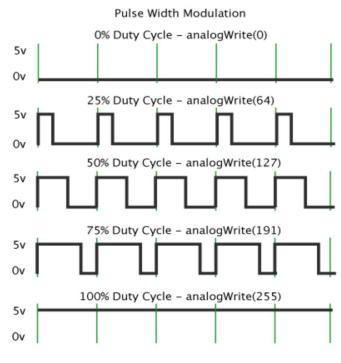


Figure 24: Pulse Width Modulation

analogWrite(led1, bri1); analogWrite(led2, bri2); analogWrite(led3, bri3); analogWrite(led4, bri4); analogWrite(led5, bri5); digitalWrite(pin3, HIGH); delay(10);

digitalWrite(pin3, LOW);

Duty Cycle: 100%, analogWrite(255)

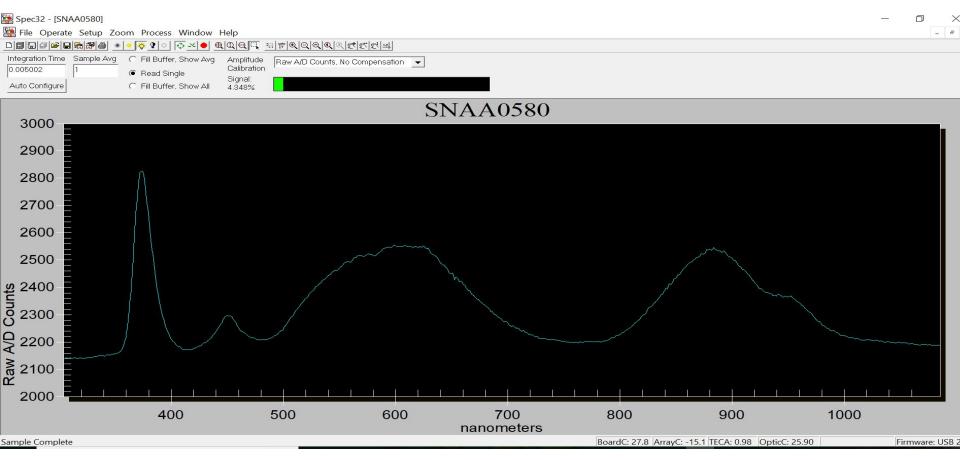


Figure 25: Spectrum at 100% duty cycle

Duty Cycle: 75%, analogWrite(191)

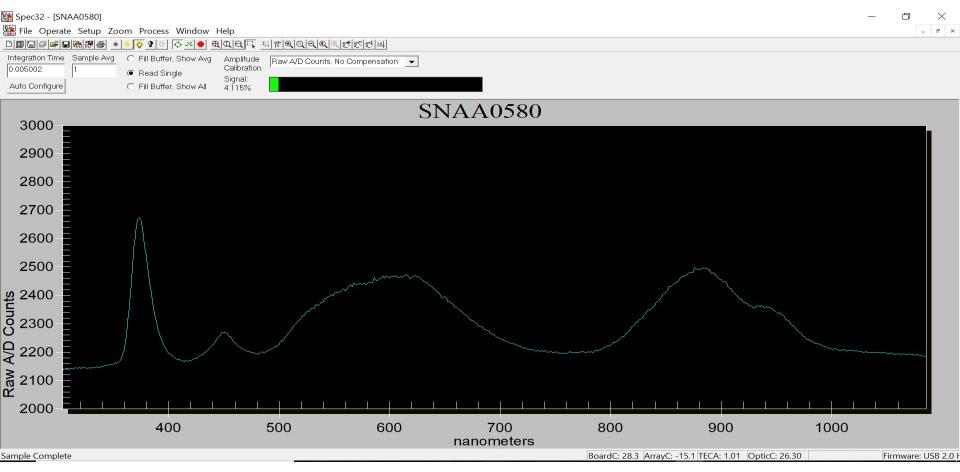


Figure 26: Spectrum at 75% duty cycle

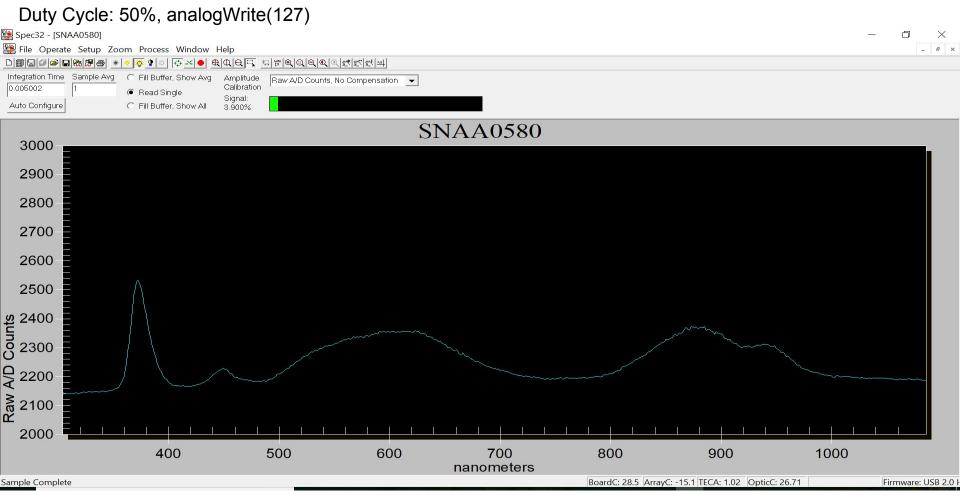


Figure 27: Spectrum at 50% duty cycle

Duty Cycle: 25%, analogWrite(64)

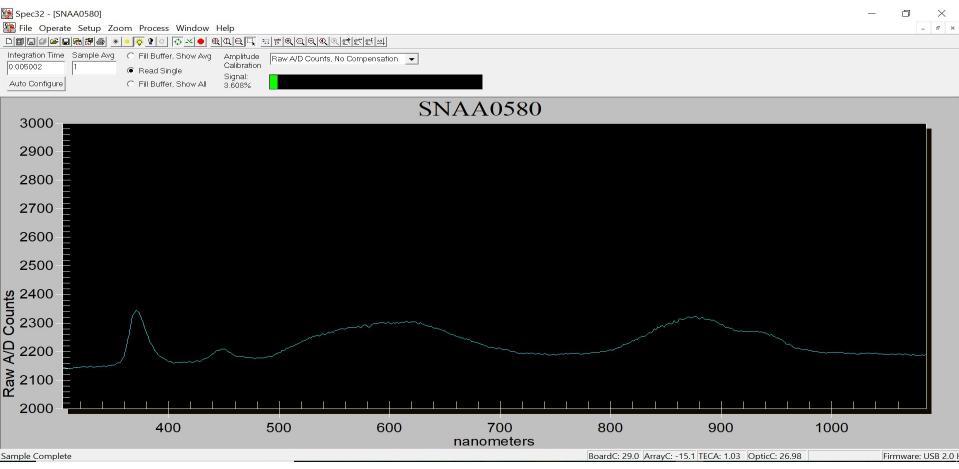


Figure 28: Spectrum at 25% duty cycle

Duty Cycle: 0%, analogWrite(0)

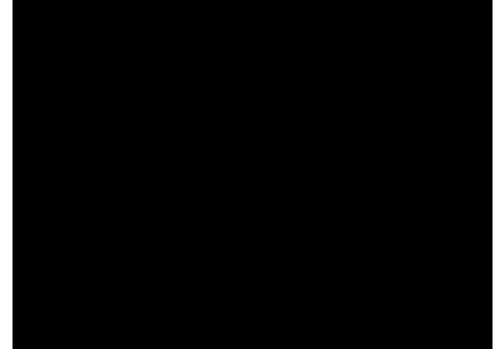
| 👑 Spec32 - [SNA 🖼 File - Operate | AA0580] e Setup Zoo | om Process Window | Help | | | | | - | - O × |
|--|------------------------|---|------------|---|-------------------|-------------------|-----------------------------|-----------|-------------------|
| | 🚓 🕼 🍯 🔹 | | 100 100 | ঽ @ @ <u>@</u> ¢ <u>@</u> { <u>@</u> | | | | | |
| Integration Time 0.005002 Auto Configure | | Fill Buffer, Show Avg Read Single Fill Buffer, Show All | | Counts, No Compensation | | | | | |
| 3000 | | | | SN | JAA0580 | | | | |
| 2900- 2800- | | | | | | | | | |
| 2700 | | | | | | | | | |
| 2600 | | | | | | | | | |
| 2500 | | | | | | | | | |
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| 2000 - | | 400 | 500 | 600 | 700 nanometers | 800 | 900 | 1000 | |
| Samp <mark>le C</mark> omplete | e | | | | | BoardC: 28.8 Arra | ayC: -15.1 TECA: 1.03 Optic | 27.12 | Firmware: USB 2.0 |

Figure 29: Spectrum at 0% duty cycle

Demonstration of putting the kernel into the light tube



Response on the spectrum as the kernel slides through the light tube



Conclusion and Future Work

Improvement on Hardware

- Circuit design to lessen heat dissipation
- Make the board thicker so that the trace can be wider (suitable for the amount of current)

Conclusion and Future Work

Improvement of Software

- Save data from spectrometer
- Improve on programming method (using Matlab GUI or Simulink in Matlab)

Credits

- John Hart
- Professor Stasiewicz and Eric Cheng
- TA Channing Philbrick
- And other TAs for this class

Reference

[1] National Cancer Institute. (2018). Aflatoxins. [online] Available at: https://www.cancer.gov/about-cancer/causes-prevention/risk/substances/aflatoxins [Accessed 19 Mar. 2018].

[2] "Aflatoxin Effect On Health", Fao.org, 2018. [Online]. Available: http://www.fao.org/fileadmin/user_upload/wa_workshop/ECAfrica-caadp/4._Aflatoxin_USAID.p df [Accessed: 05- Feb- 2018].

[3] T. C. Pearson, D. T. Wicklow, E. B. Maghirang, F. Xie and F. E. Dowell, "DETECTING AFLATOXIN IN SINGLE CORN KERNELS BY TRANSMITTANCE AND REFLECTANCE SPECTROSCOPY", Transactions of the ASAE, vol. 44, no. 5, 2001.

[4] "Detection of Aflatoxin Contaminated Figs Using Near-Infrared (NIR) Reflectance Spectroscopy", Research Gate, 2018. [Online]. Available: https://www.researchgate.net/publication/262684486_Detection_of_Aflatoxin_Contaminated_Fig s_Using_Near-Infrared_NIR_Reflectance_Spectroscopy [Accessed: 05- Feb- 2018].

[5] A. Wacoo, D. Wendiro, P. Vuzi and J. Hawumba, "Methods for Detection of Aflatoxins in Agricultural Food Crops", Hindawi, 2018. [Online]. Available: https://www.hindawi.com/journals/jac/2014/706291/ [Accessed: 03- Feb- 2018].

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